

# Proteins in Materials Science

November 13<sup>th</sup>, 2004



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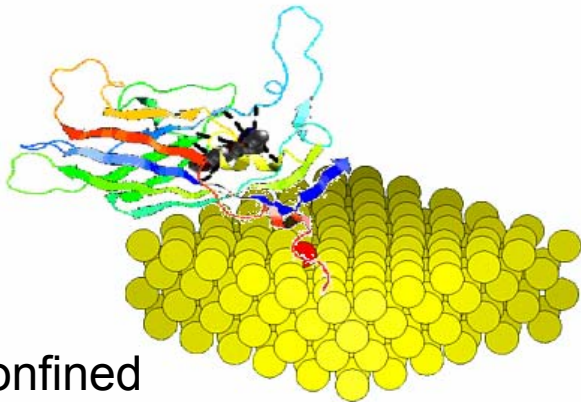


# Protein Theory and Simulations

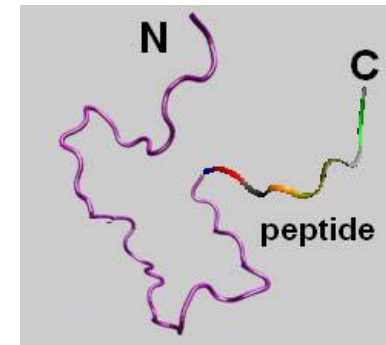
Protein modeling : Analytical approaches (GNM), chemical kinetic equations, molecular simulations (AMBER, XMD), ab initio calculations (VASP) (in collaboration with Materials Research Institute, Penn State)



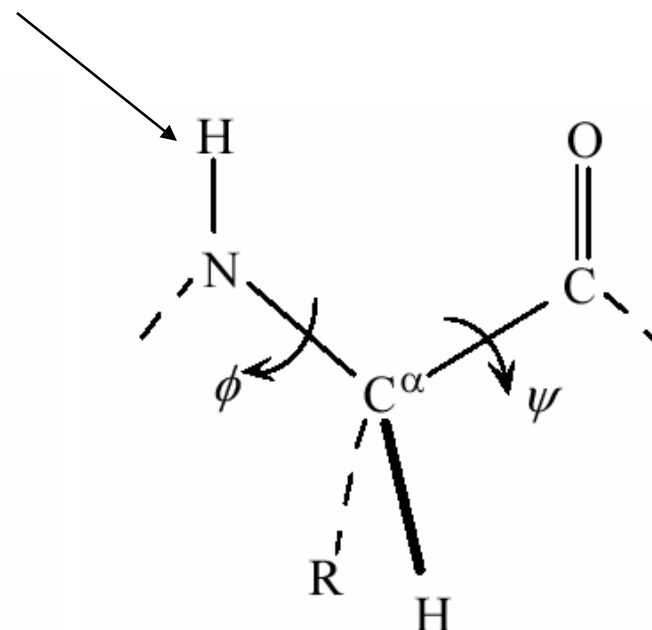
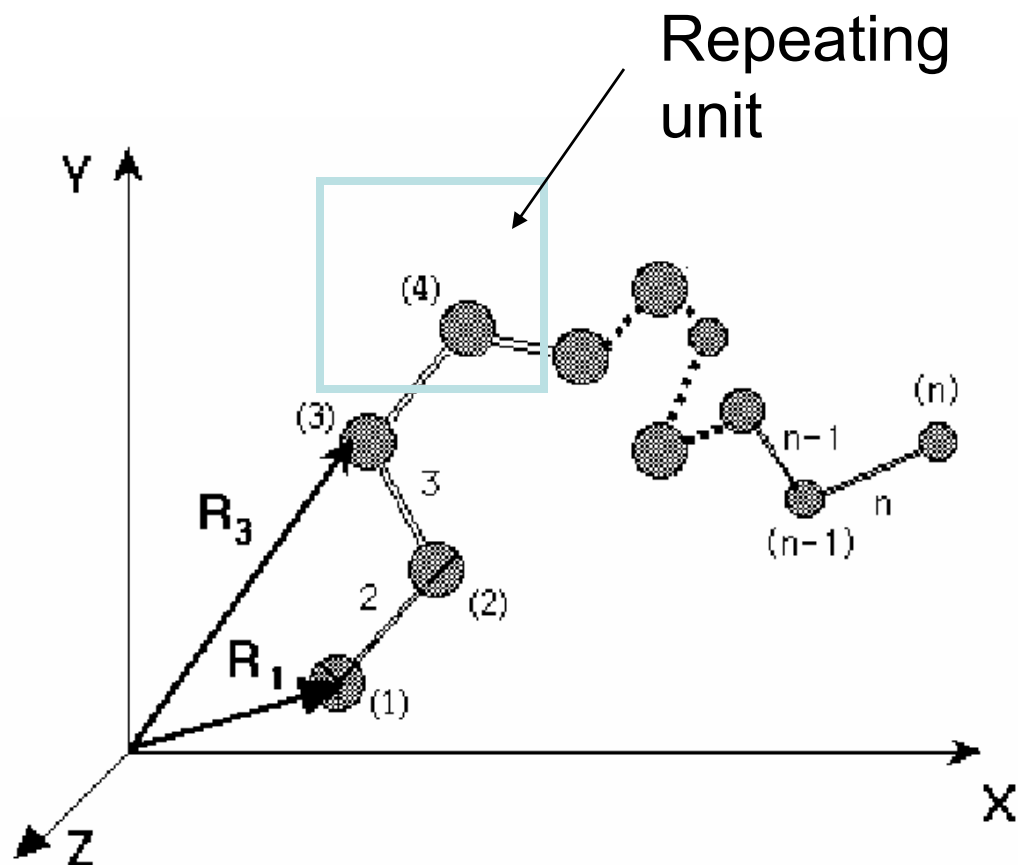
LION-XM  
128 node super  
computers (cost  
½ million \$)



Confined  
Environments:  
Protein-Surface  
Interactions



Protein Design



Proteins are heteropolymers made of amino acids

# MOLECULAR DYNAMICS SIMULATIONS

- A deterministic method based on the solution of Newton's equation of motion

$$\mathbf{F}_i = m_i \mathbf{a}_i$$

for the  $i$ th particle; the acceleration at each step is calculated from the negative gradient of the overall potential, using

$$\mathbf{F}_i = - \text{grad } V_i = - \nabla V_i$$

## LIMITATIONS OF MD

- Full atomic representation  $\rightarrow$  noise
- Empirical force fields  $\rightarrow$  limited by the accuracy of the potentials
- Time steps constrained by the fastest motion (bond stretching of the order of femtoseconds)
- Inefficient sampling of the complete space of conformations
- High computational cost: Limited to small proteins (100s of residues) and short times (subnanoseconds)

## *ALTERNATIVE METHOD* *COARSE GRAIN SIMULATIONS*

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## GAUSSIAN NETWORK MODEL (GNM)

Demirel, M.C. (with others)  
Protein Science,  
December 1998

$$\mathcal{H} = \frac{1}{2} \gamma [\Delta \mathbf{R}^T (\Gamma \otimes \mathbf{E}) \Delta \mathbf{R}],$$

$$\Gamma^{-1} = (\gamma / 3k_B T)^* \langle \Delta \mathbf{R} \Delta \mathbf{R}^T \rangle$$

$$A = -k_B T \ln Z_N = -(3k_B T / 2) \ln [(\pi / \gamma^*)^{N-1} \det(\Gamma^{-1})],$$

$\Gamma$ : Connectivity matrix,  $\Delta \mathbf{R}$ : fluctuation of each residue

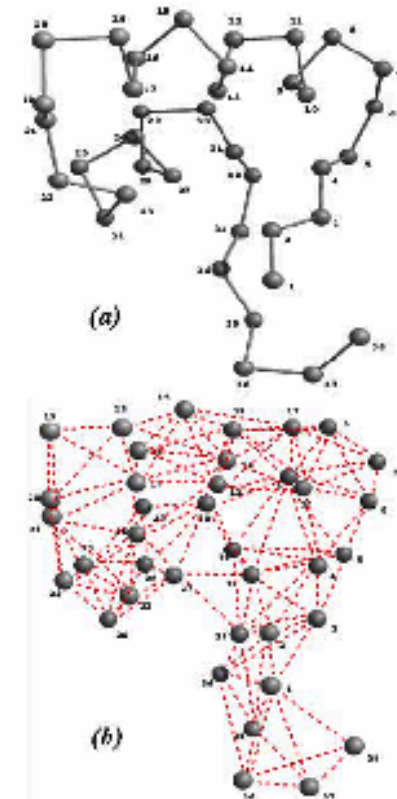
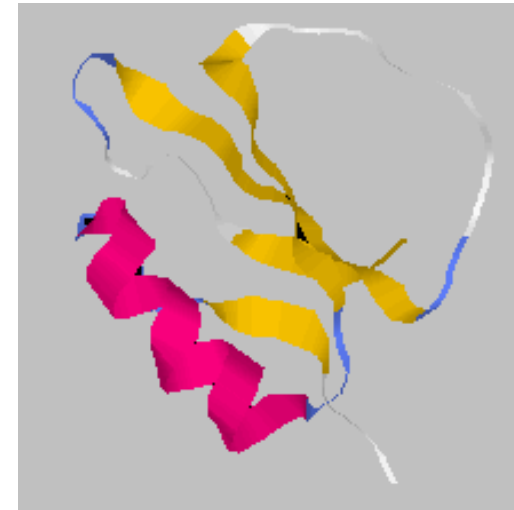
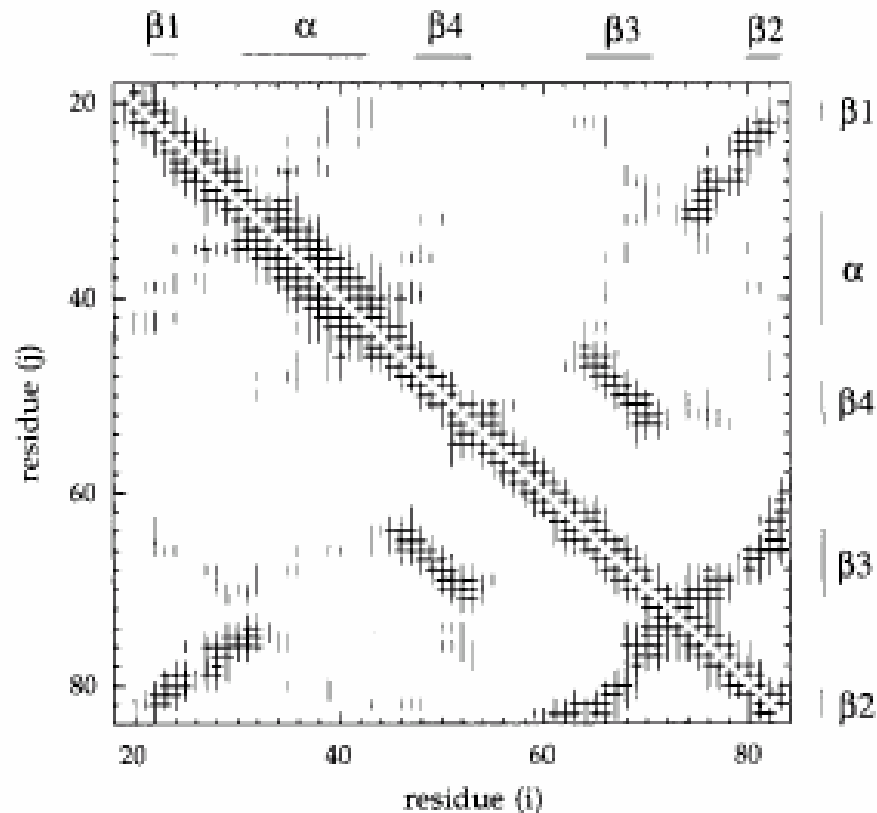


Fig 1. GNM of biomolecules.  
The set of representative  
interaction sites in (a) forms the  
nodes of the network in (b)

# CONTACT MAP (CONNECTIVITY MATRIX)



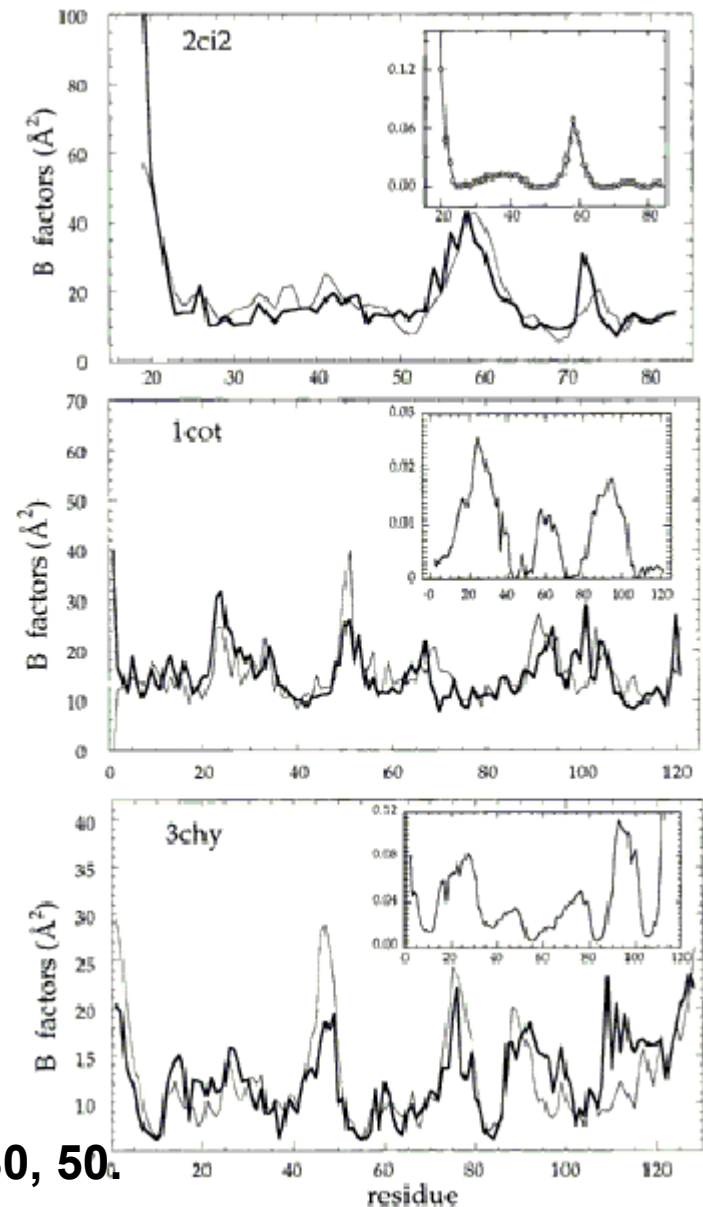
Chymotrypsin  
inhibitor-2 64 residues

$$\Gamma_{ij} = \begin{cases} -H(r_c - r_{ij}), & i \neq j \\ -\sum_{i(\neq j)}^N \Gamma_{ij}, & i = j \end{cases}.$$

Demirel et al. (1998), Protein Science, v7, 2522

# COMPARISON

... of theoretical (thick curve) and experimental (thin curve) B factors for Chymotrypsin inhibitor 2 (2ci2), C2 protein (1cot), and CHE-Y protein (3chy)



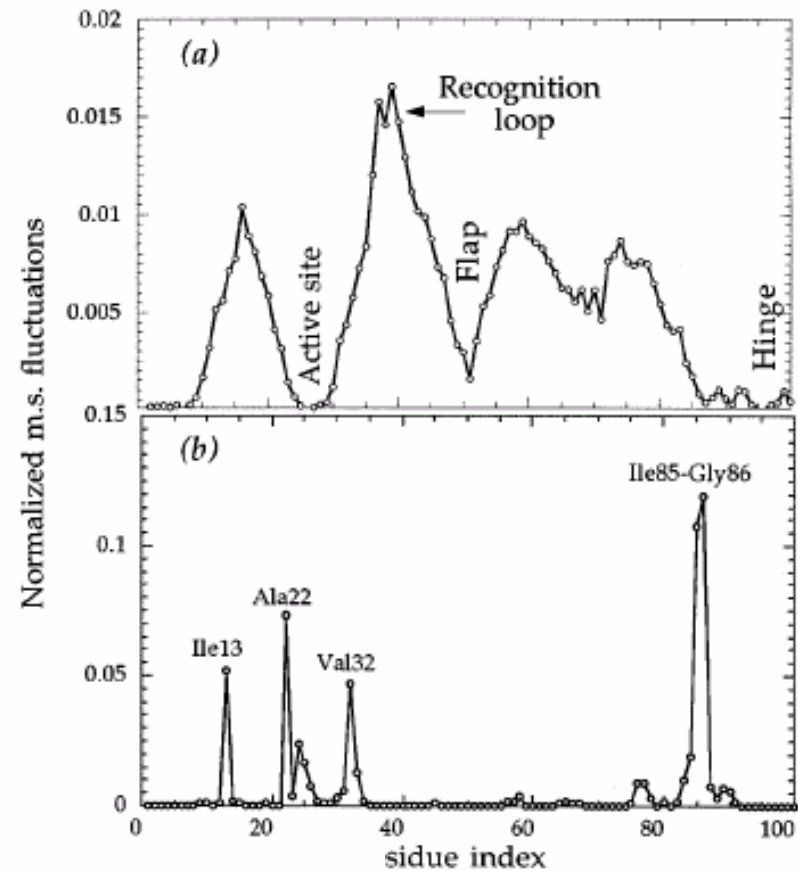
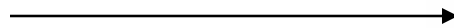
Atilgan, et al., 2001, Biophysical Journal, v80, 50.



# SLOW AND FAST MOTION FUNCTION AND STABILITY

$$\Gamma^{-1} = (\gamma/3k_B T) \langle \Delta \mathbf{R} \Delta \mathbf{R}^T \rangle$$

HIV-1 protease

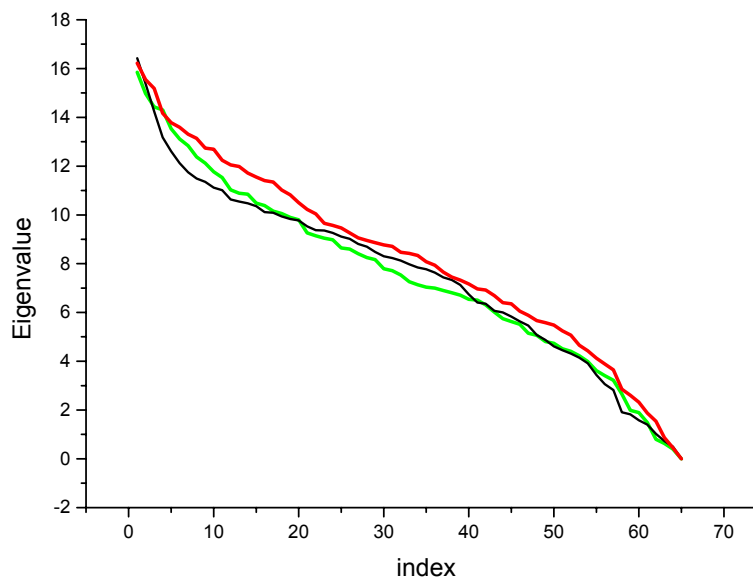
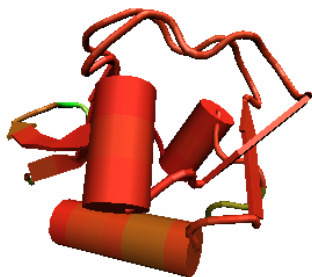
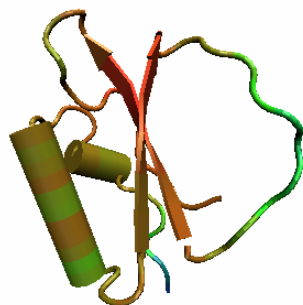


$$\begin{aligned} \langle \Delta \mathbf{R}_i \cdot \Delta \mathbf{R}_i \rangle_k &= (3k_B T / \gamma) [\lambda_k^{-1} \mathbf{u}_k \mathbf{u}_k^T]_{ii} \\ &= (3k_B T / \gamma) \lambda_k^{-1} [\mathbf{u}_k]_i [\mathbf{u}_k]_i, \end{aligned}$$

Bahar, et al., (1998), Physical Review Letters, v80, 2733



# Eigenvalue Distribution

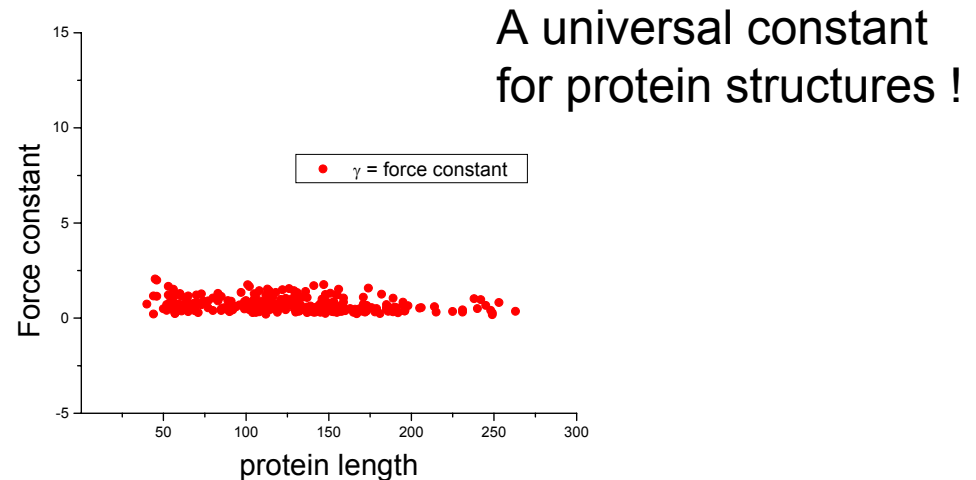
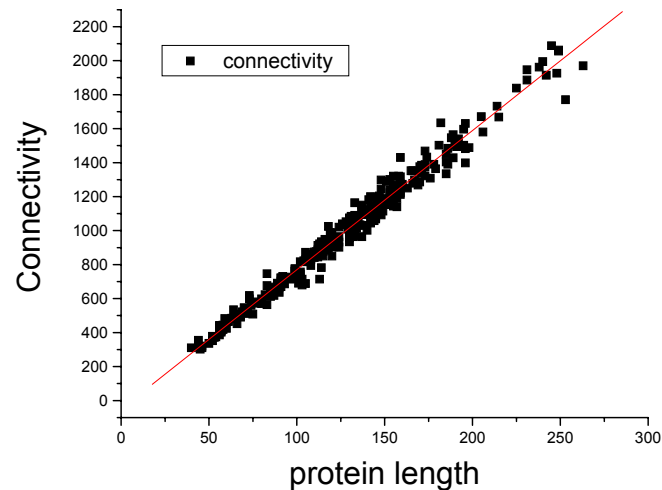


$$connectivity \equiv \sum_i \Gamma_{ii} = \sum_k \lambda_k$$

Eigenvalue distributions are similar for proteins which have equal length

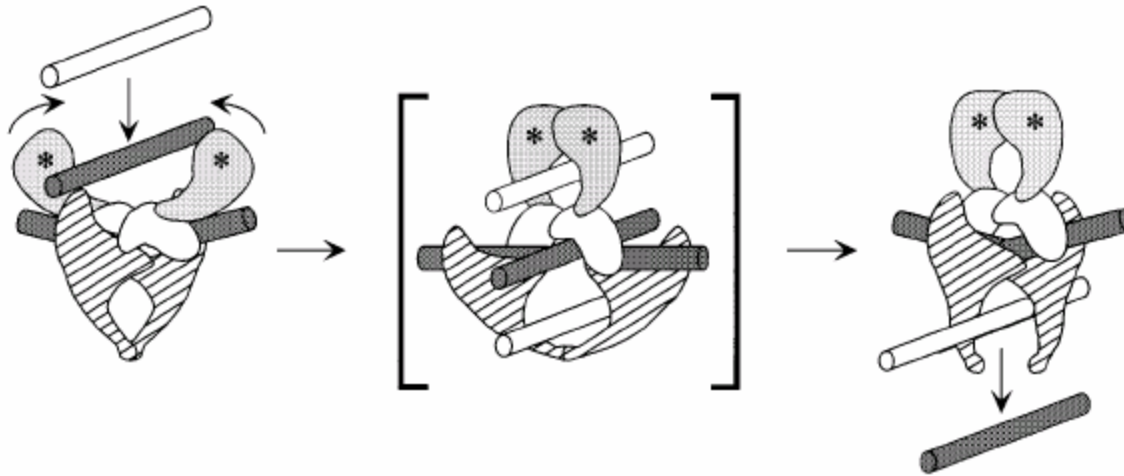
Length=65

# Connectivity and Force constant



532 protein X-ray crystallography data which is extracted from PDB:  
connectivity is linearly varying with the protein length and  $\gamma$  is a  
constant ( $\sim 1$  kcal/mol  $\text{\AA}^2$ )

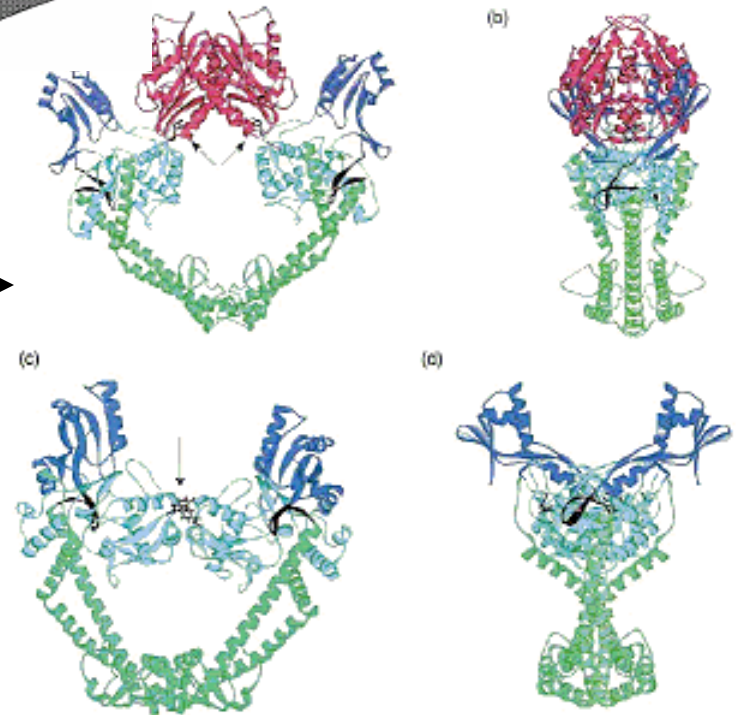
# FUNCTION



DNA topo II and GyrA  
crystallographically determined fragments  
shown in two different views each.

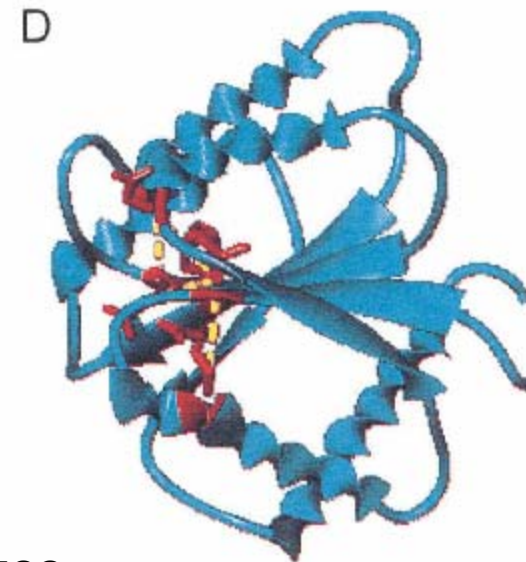
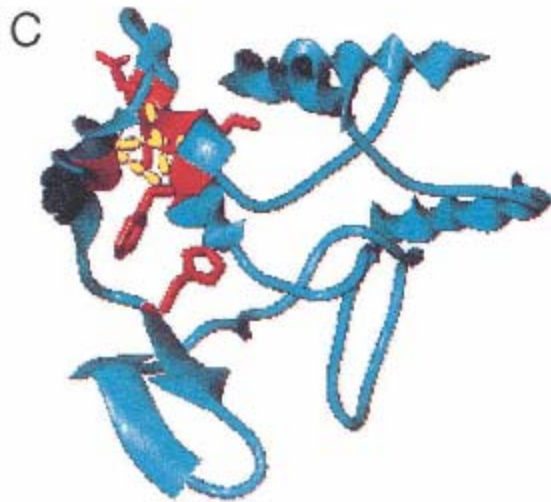
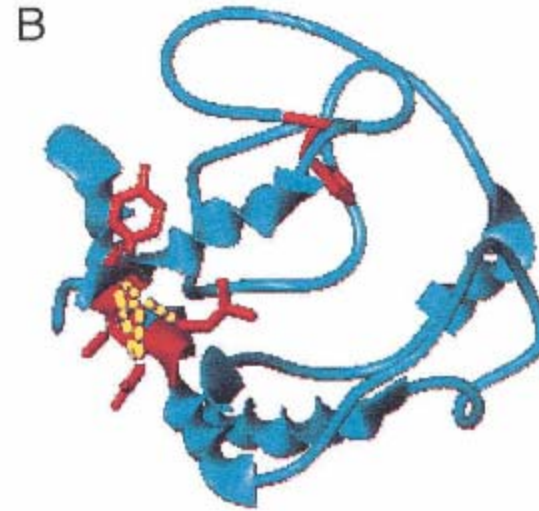
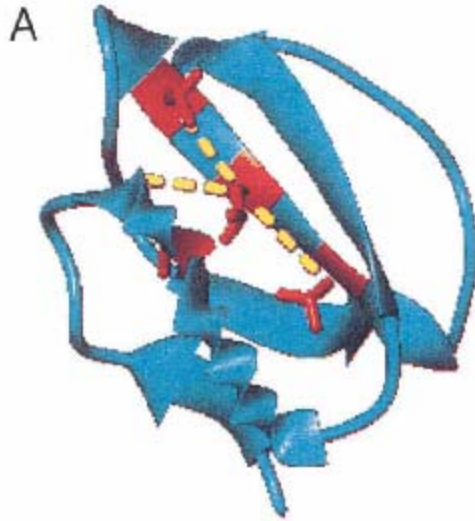
Colors denote the functional motions  
obtained from GNM **mode analysis**

Demirel (with others) 1999, Int. J. Quantum  
Chem., Vol. 75,, pp. 301



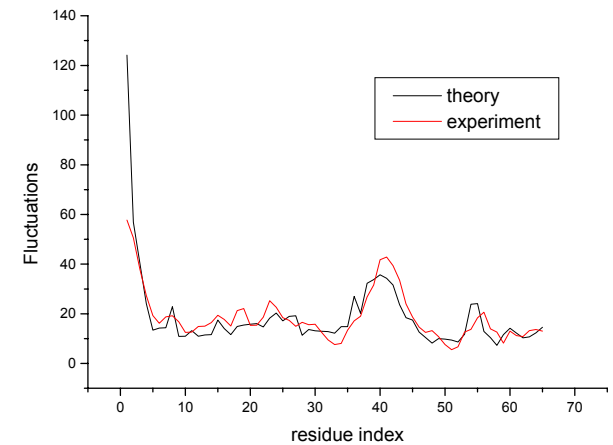
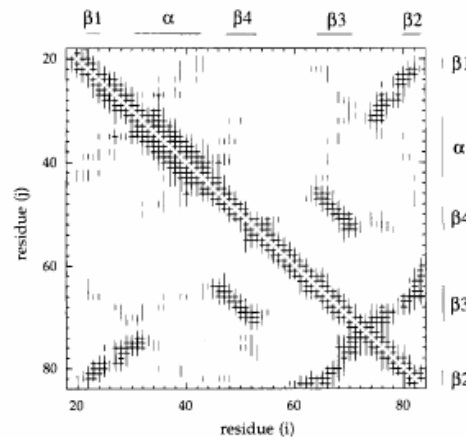
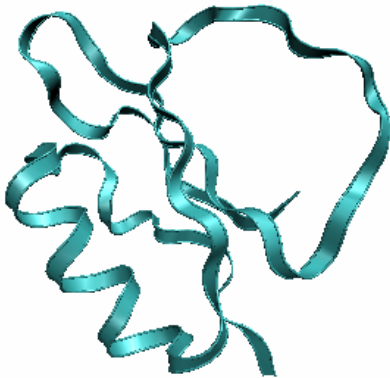
# STABILITY

Fast modes  
determine  
stability

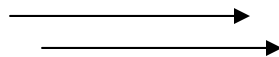


Demirel et al. (1998), Protein Science, v7, 2522

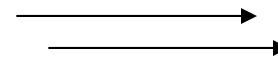
# Elastic Network Model: Summary



Structure



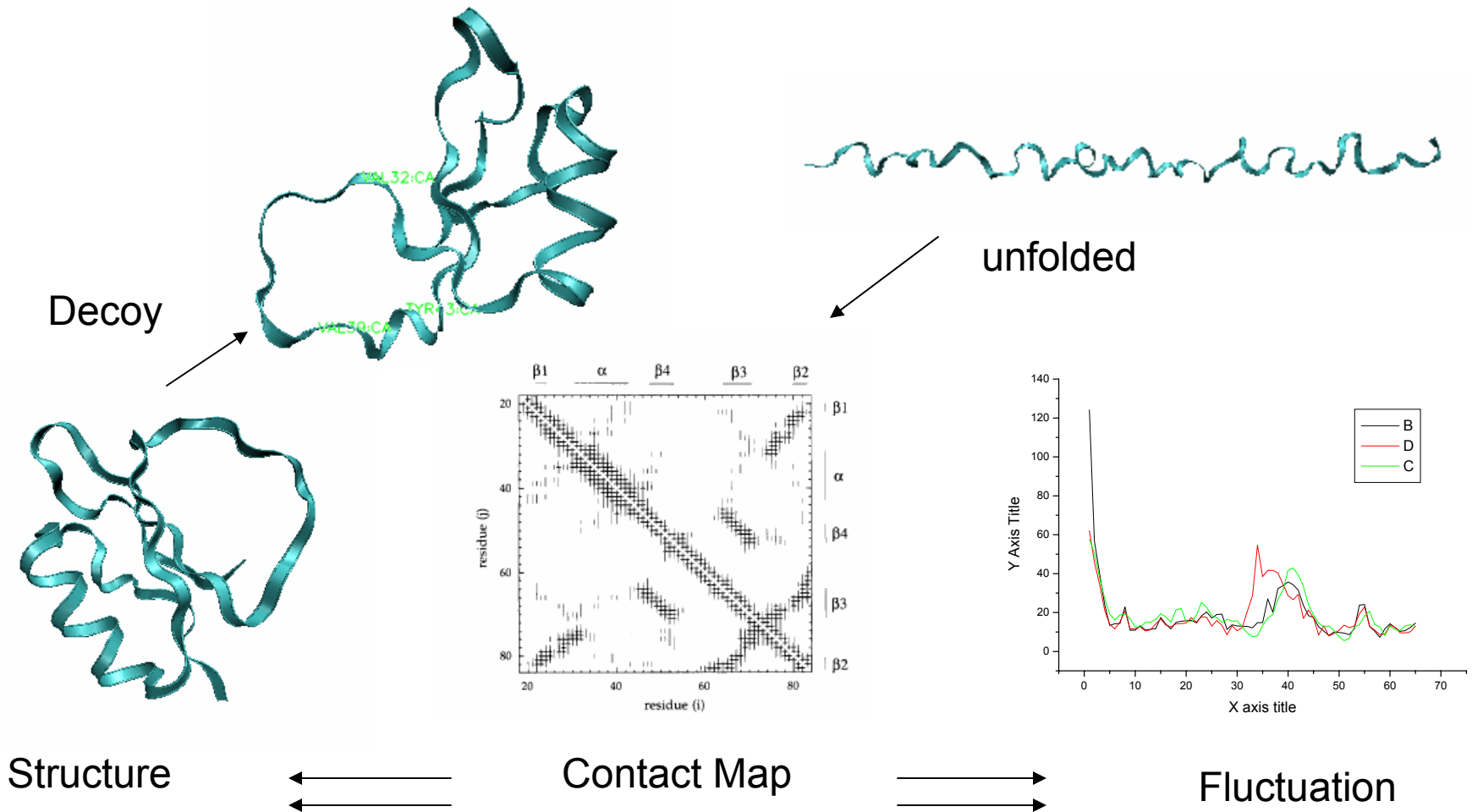
Contact Map



Fluctuation

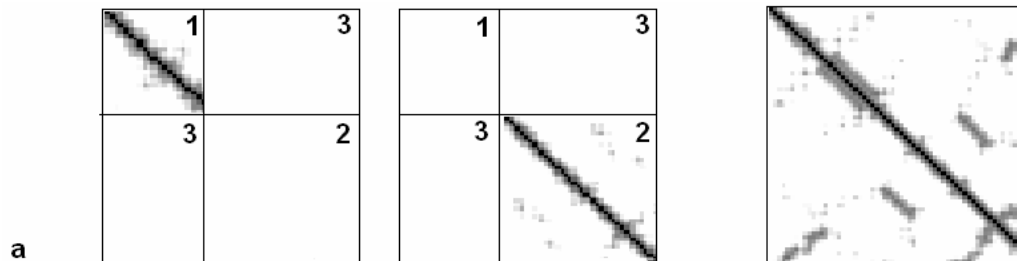
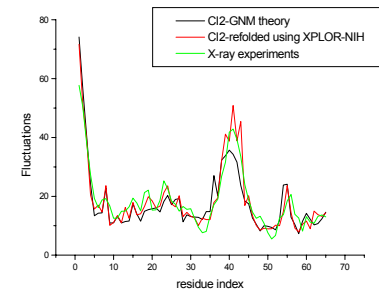
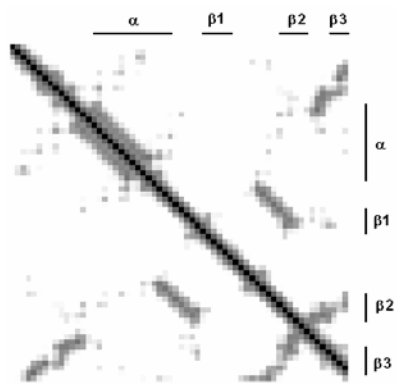


Function, Stability

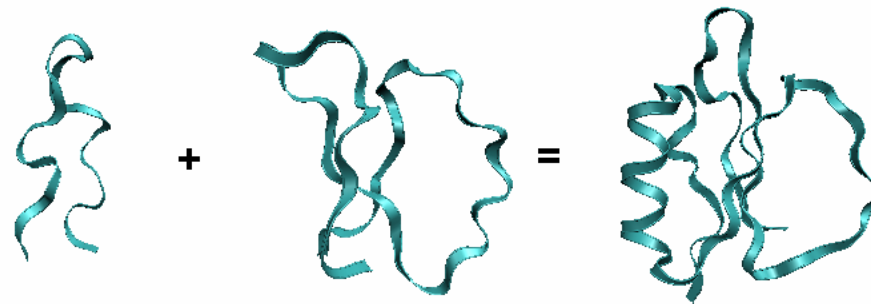


Demirel, M.C., Cherny, D., **Clustering and Diversity of Surface Fluctuations for Proteins**, submitted, 2004

# “BUILDING BLOCKS” of PROTEINS

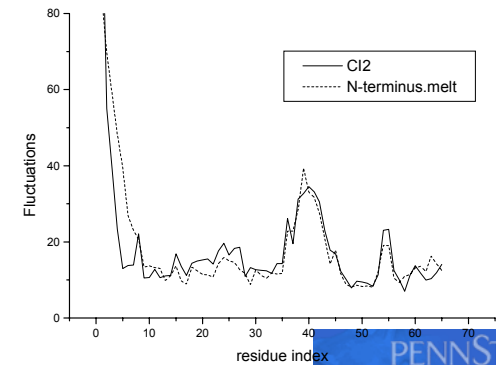
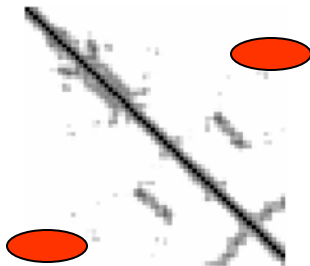
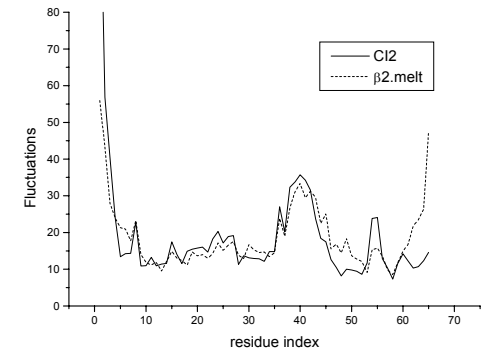
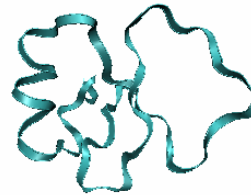
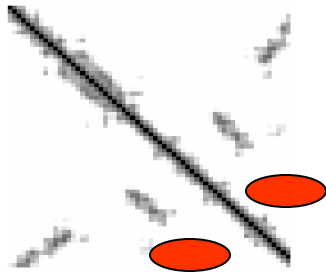
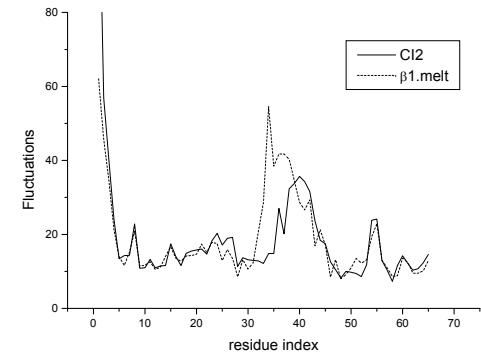
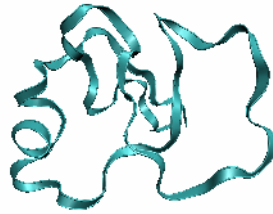
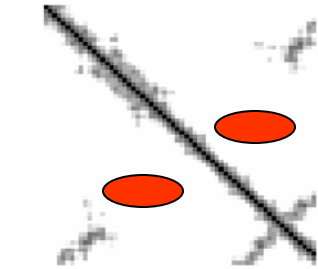


a



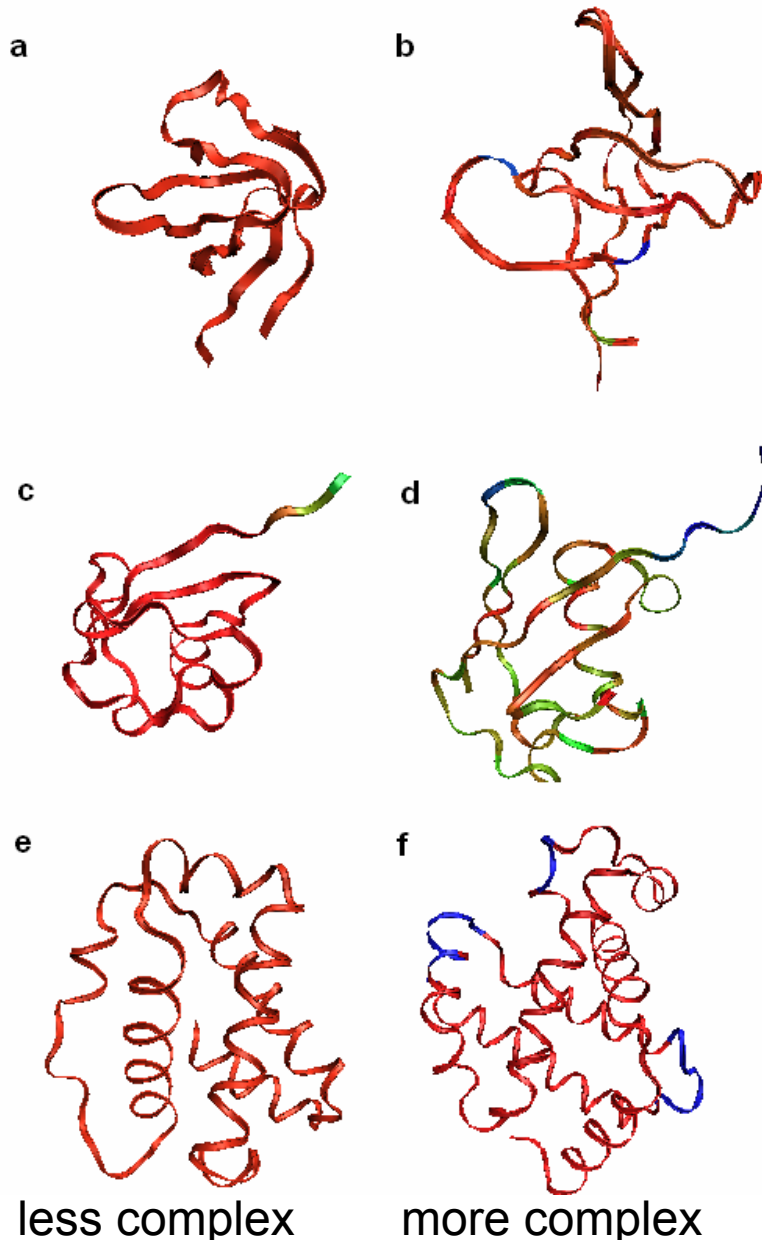
b

# CLUSTERING OF FLUCTUATIONS





# PROTEIN COMPLEXITY FROM FLUCTUATIONS



The highly flexible regions are shown in blue and the least flexible with red.

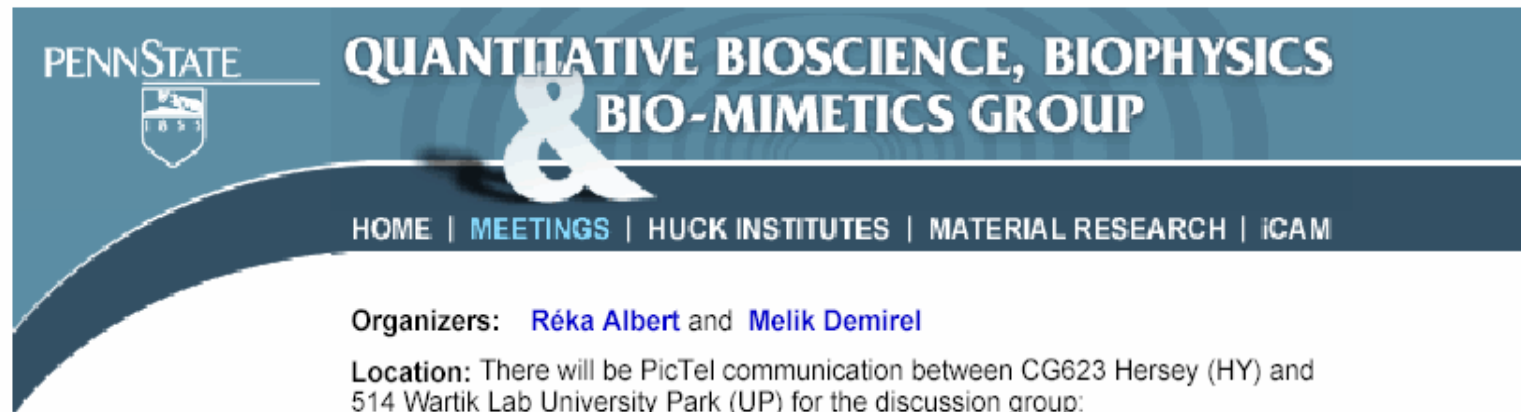
SH3 domains are from nematode *C. elegans* (pdb.3sem) (a) and *H. sapiens* (pdb.5hck) (b) ; ubiquitin is from *E. coli* (pdb.1f0z) (c) and *H. sapiens* (pdb.1ubi) (d); and hemoglobin is from *Paramecium* (pdb.1dlw) (e) and *H. sapiens* subunit (pdb.1bz0, chain B) (f).

Demirel, M.C., Keskin, O., **Protein Interactions and Fluctuations in a Proteomic Network using an Elastic Network Model**, Vol 22, JBSD, 2004

# Conclusion

- Elastic network models are successful in describing equilibrium protein motions.
  - ***Experimental results (X-ray, NMR relaxation) agree well*** with this simple model.
  - Our approach introduces a new concept for classifying ***building blocks of proteins*** based on thermal fluctuations.
  - ***Large biological assemblies*** (viruses, structural proteins-titin, biomachinaries) can be simulated using elastic network models
- Protein Networks will open new avenues for biology / biophysics applications. Understanding binding of proteins will improve our conceptions about “how nature design proteins”.

# Outreach: Quantitative Bioscience seminars at Penn State



**Organizers:** [Réka Albert](#) and [Melik Demirel](#)

**Location:** There will be PicTel communication between CG623 Hersey (HY) and 514 Wartik Lab University Park (UP) for the discussion group:

**Time:** 1:00pm-2:00pm Wednesday (see the agenda below)

**How to join:** If you are interested, please e-mail me ([mcd18@psu.edu](mailto:mcd18@psu.edu)) so that we will add you to the list.

Click to the links on agenda for extra information and presentations (pdf)

## AGENDA:

**FALL'2003**

**19-November-2003**

- Welcome and discussion: Reka Albert & Melik Demirel [welcome.pdf](#)

# ACKNOWLEDGEMENT

- **GROUP**

- Murat Cetinkaya (Ph.D. Student)
- Eric So (M.S. Student)
- Lee Salway (Undergrad-EE)

- **COLLABORATORS**

- ***Protein Simulations***

O. Keskin: National Cancer Institute,  
National Institutes of Health, MD, USA  
I. Bahar: University of Pittsburgh Medical  
School, PA, USA  
J. Sofo: Pennsylvania State University,  
Physics, PA, USA

- ***Green Fluorescence Protein***

A. Zeytun, A. Bradbury: Los Alamos  
National Laboratory, NM, USA

- ***Protein Assembly & Q-dots***

T. Jovin: Max Planck Institute for Biophysical  
Chemistry, Göttingen, Germany  
K.Kalkan: Nanofab, Pennsylvania State  
University, PA, USA  
O. Mayans: Biozentrum, University of Basel,  
Basel, Switzerland

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- National Science  
Foundation-ICAM funding



- Penn State University,  
MRSEC-MRI/Huck Seed  
Grant

- Penn State University,  
Start-up funds



- Alexander von Humboldt  
Fellowship

